Larval fishes identification

Laboratory procedure

Zooplankton biomass measurement (Displacement or Settled volume)

Sorting

Fish eggs

Fish larvae

Identify

Count

Size Measurement

Standardization

Sorting

1. Draining the samples through a hand net (mesh size same as plankton net). Use two jugs when draining
2. Return the fixatives into an original sampling jar
3. Washing the specimens with fresh water
4. Retain specimens in a jug with fresh water
5. Stir the solution softly with glass rod
6. Pour a small amount of samples into a petri-dish.
7. Use tiny thin stainless steel forceps to pick up fish eggs and larvae from the petri-dish under dissecting microscope at a magnification of about 10 times (10x ocular, 1x objective) and place in a label petri-dish. Precaution must be taken to prevent deterioration of specimens when picking. Count the number of fish eggs and larvae by using counter to record the number while you are removing them from the sample. (During sorting you may preliminary identify your specimens and count and record the number)
8. Keep doing 6 and 7 a little bit by bit until no more sample left
9. Store fish eggs and larvae into separate labeled vial. The preservative fluid in the vial is 70% ethanol. Place label in each vial, the information on label including sampling date, station number, sampling site, sampling method (oblique or horizontal or vertical tow), plankton net type. Use pencil for labeling
10. In case that you cannot finish sorting by one day. Please put back the unsorted plankton into 5% buffer formalin. Then continue sorting with the above process on the next day
11. After sorting, the remaining plankton will be replaced in the original sampling jar for processing later.
12. The total number of fish eggs and larvae removed from the sample are recorded on the plankton work sheet